ISSN PRINT 2319 1775 Online 2320 7876

Research Paper © 2012 IJFANS. All Rights Reserved Journal Volume 13, Iss 4, 2024

PHYSICO-CHEMICAL PARAMETERS AND DISTRIBUTION OF PHYTOPLANKTON IN GUNDLA CHERUVU IN ARMOOR VILLAGE, ARMOOR MANDAL, NIZAMABAD DISTRICT, TELANGANA STATE.

A. Pradeep Kumar, Prof. Nirmala Babu Rao

Department of Botany, University College of Science, Osmania University, Hyderabad, Telangana, India.

ABSTRACT

From November 2014 to October 2015, a study was conducted on Gundla cheruvu in Armoor village, Armoor mandal, Nizamabad District, Telangana state. Samples were collected in the morning each month to analyze their physical, chemical, and biological characteristics. The examined physico-chemical parameters included Water Temperature, Turbidity, pH, Biological Oxygen Demand (BOD), Chemical Oxygen Demand (COD), Dissolved Oxygen (DO), Total Dissolved Solids (TDS), Total Suspended Solids (TSS), and nutrients such as Chloride, Nitrate, Phosphate, Sulphates, and Silicates. Results indicated that the concentration of physico-chemical parameters was higher in the summer than in other seasons. The primary producer community, known as phytoplankton diversity, consisted of various algae forms, which serve as significant biological indicators of water quality. The study suggested that Gundla cheruvu was at risk of eutrophication, emphasizing a potential combination of water quality indices and pollution contamination.

Keywords

Physico-chemical analysis, Phytoplankton diversity, Gundla cheruvu.

INTRODUCTION

Water is an essential component of all living organisms and plays a crucial role in the biosphere, biogeochemical cycles, and Earth's ecosystem. Freshwater resources are increasingly scarce, and the rapid decline in water quality has become a national concern, Abdar (2013). Threats to global freshwater biodiversity can be categorized into five interacting groups: overexploitation, water pollution, and habitat degradation, Agarwal (1999), Abd-Ellatif et al., (2016). To prevent the use of contaminated water for human consumption, it is crucial to thoroughly understand the ecosystems of this lake. Local populations use lake water for various purposes, including bathing, laundry, and washing kitchenware. Additionally, inlets discharging water into lakes often carry chemical pesticides and fertilizers, further contaminating the water. Considering the environmental impact of water chemistry, biological assessment is a valuable alternative for evaluating the ecological quality of aquatic ecosystems, Amin Hossaini Motlagh, Navatha and Manikya Reddy (2013).

Eutrophication is a phenomenon linked to the nutrient enrichment of aquatic ecosystems, resulting from the gradual aging process of lakes. This process, known as succession, is



ISSN PRINT 2319 1775 Online 2320 7876

Research Paper © 2012 IJFANS. All Rights Reserved Journal Volume 13, Iss 4, 2024

exacerbated by the inability of lakes to purify themselves, making them prone to accumulating pollutants and becoming more intricate and delicate. The decline in water quality and the shrinkage of water bodies, driven by increasing anthropogenic influence in the catchment area, have accelerated eutrophication. Understanding the distribution of plankton and the stratification of lakes requires consideration of physical characteristics such as temperature, light penetration, and water movement. These factors collectively determine the lake biological community and consequently, the quality of the water. Both human activities and natural dynamics can significantly alter a lake physico-chemical characteristics, impacting the quantity and quality of the water, the distribution and diversity of species, the lake productive capacity, and the balance of the surrounding ecological system.

Phytoplankton form the foundation of nutritional cycles in aquatic ecosystems. Their composition is influenced by numerous factors that change with ecological shifts. The structure of the ecosystem relies heavily on phytoplankton and the composition of their communities, Abd-Ellatif et al., (2016). As primary producers, phytoplankton are a vital energy source and the building blocks for all life in aquatic environments. Photosynthetic primary production ultimately drives productivity at higher trophic levels. Eutrophication has already pushed some lakes to the brink of disappearance. Therefore, to prevent further degradation, these lakes need focused attention and a thorough understanding of their ecosystems. Water Quality Indices vary, while pollution indices calculations indicate that these lakes are contaminated and at risk of eutrophication.

MATERIALS AND METHODS

The majority of the analysis, including measurements of Water Temperature, Turbidity, pH, Biological Oxygen Demand (BOD), Chemical Oxygen Demand (COD), Dissolved Oxygen (DO), Total Dissolved Solids (TDS), Total Suspended Solids (TSS), and concentrations of Chloride, Nitrate, Phosphate, Sulphates, and Silicates is conducted through laboratory tests as outlined by Altaf Ganai and Saltanat Parveen (2014).

Planktonic algal species were sampled monthly from Gundla cheruvu between November 2014 to October 2015. Terrestrial algae were collected early in the morning using a knife, while benthic algae were retrieved with forceps. Samples were preserved in a 4% formaldehyde aqueous solution in the lab for research purposes. Morphological examinations of the fresh material were conducted using a light microscope. The identification of taxa was based on the methodologies of Airill, Querijero and Ching (2016), Arumugam, Sivakami and Premkishore (2015), Tiwari, Rana, and Chauhan (2006), Bowling (2009) and Boyd et al. (2013).

RESULTS AND DISCUSSION

Physico-chemical parameters

The temperature range at Gundla cheruvu varied from 21.30 to 32.50°C shown in Table-1 and Figure-1, Similar reports on temperature of aquatic ecosystems were produced by Ross (1969),



ISSN PRINT 2319 1775 Online 2320 7876

Research Paper © 2012 IJFANS. All Rights Reserved Journal Volume 13, Iss 4, 2024

Thomas (1973), Swarnalatha and Narasingrao (1998), Bhalla et al., (2006) and Chinnaiah et al., (2011). Turbidity values ranged from 19.4 to 29.5 at Gundla cheruvu, Table-1 and Figure-1, Comparable observations have been made by Krishnamoorthi et al., (2011), Young et al., (1985), Garg et al., (2006), and Solanki et al., (2015).

A pH meter scale measures the concentration of hydrogen ions in a solution, indicating its acidity or alkalinity. pH fluctuations in water can affect various physicochemical properties. The pH values ranged over a year at Gundla cheruvu from 6.9 to 8.9, With alkaline conditions promoting the growth of primary producers, consistent with findings reported by Ishaq & Khan (2013), Kumar and Prabhahar (2012). The estimated BOD content as shown in Table-1 and Figure-1. In November 2014 to October 2015, The BOD range at Gundla cheruvu 8.2 to 17.2. In findings, Mittal and Senegar (1989) and Warwick and Wolverton et al., (1983) noted reduced BOD readings throughout the winter and attributed this to phyto populations as the main cause. Blue greens were observed by Panday and Tripati (1984) to be both qualitatively frequent and directly connected to changes in BOD, Table-1 and Figure-2.

The range in COD values are shown in Table-1 and Figure-2. The COD values range for a year at Gundla cheruvu 83 to 161. In their studies on the ecology of water bodies, Trivedy et al., (1990). increased the use of BOD and COD to assess the standards of water quality. Monthly COD was high in the summer, which was consistent with Chatterjee (1992) observations. Sinha and Biswas (2016) and Rajashekar et al., (2010), recently completed comparable work in the field of water bodies. The DO range for a year at Gundla cheruvu 5.8 to 10.6 shown in Table-1 and Figure-2. A higher DO value in an aquatic environment indicates more photosynthetic activity and less organic matter that will be destroyed by biological processes. These findings concurred with those of Kannel et al., (2007), Rai (1978), Shashikant and Raina (1990), Rajakumar and Ramanibai (1994).

The TDS range for a year at Gundla cheruvu 122 to 240 shown in Table-2 and Figure-3, Hascoet et al., (1986), Subba Rao (1981), are a few notable reports that are comparable to the current study. The total dissolved solids are unevenly distributed throughout the many water bodies in India, Chinnaiah et al., (2011), Sharma et al., (2010), Kumbhar et al., (2009). The TSS values range for a year at Gundla cheruvu 85 to 506 shown in Table-2 and Figure-3. TSS is a benchmark that protects the supporting food web of lakes, Berry et al., (2003), Chapman et al., (2017), Lloyd (1985), Arruda et al., (1983), Caux et al., (1997).

The Chloride content range for a year at Gundla cheruvu 77 to 97.7 shown in Table-2 and Figure-3. Munawar 1970, a high chloride content suggests an increased level of organic contamination. The findings of the content of chlorides has a correlation with the purity or impurity of water. The nitrates ranged from 0.42 to 0.92 shown in Table-2 and Figure-4. According to Jones et al., (1993), nitrate in surface water is a significant element in determining the quality of the water, with waste discharges and synthetic nitrogenous fertilizers being the



ISSN PRINT 2319 1775 Online 2320 7876

Research Paper © 2012 IJFANS. All Rights Reserved Journal Volume 13, Iss 4, 2024

main contributors. However, bacterial oxidation and plant nitrogen fixation also make a small but significant contribution, EPA (2001). According to Mithani et al., (2012), the increased levels of nitrate during the monsoon season were mostly caused by the nitrate to rich runoff from agricultural areas and a significant amount of sewage that was tainted. Khound et al., (2012), Lalparmawii (2007), Singh, M.R. and Gupta, A. (2010) all observed similar findings.

Table-2 and Figure-4, illustrate the range of phosphate concentrations found in the sampling site. For a year at Gundla cheruvu 0.13 to 0.90. The research cites previous studies by Mithani et al., (2012), Dwivedi and Pandey (2002), Jones et al., (1993), Lalparmawii (2007), Singh and Gupta (2010), and Khound et al., (2012) to support the significance of nitrate levels in surface water quality assessment. The results of sulfate ranged for a year at Gundla cheruvu 19.5 to 40.1 shown in Table-2 and Figure-5. Sulfate plays a crucial role in the aquatic environment due to its various effects. It can act as a lethal agent against slugs and snails and help regulate algae populations. Moreover, sulfate is essential for the aquatic environment, as it aids in the treatment of bacterial and ectoparasitic disorders. Shuhaimi et al., (2010), Saifullah et al., (2014), and Wani et al., (2013) have all contributed to understanding the significance of sulfate in aquatic ecosystems through their research. The silicate content in the water samples collected from Gundla cheruvu displayed a distinct trend in the range in silicate content, which is a common component of the water found in any body of water. During the year of November 2014 to October 2015, at Gundla cheruvu 2.1 to 5.2 shown in Table-2 and Figure-5, Das and Pandey(1978), Garg et al., (2006 and 2009) presented reports indicating the impact of silicates in relation to pollution of water body.

Table-1: Physico-chemical parameters of Gundla cheruvu showing ranges expressed in (mg/L), except Temperature(°C), Turbidity (NTU) and pH for the year 2014-2015

Rang e	Temperature	Turbidit y	рН	Biological Oxygen Demand	Chemical Oxygen Demand	Dissolved Oxygen
Min	21.3	19.4	6.9	8.2	83	5.8
Max	32.5	29.5	8.9	17.2	161	10.6

Note: Max=Maximum, Min=Minimum



ISSN PRINT 2319 1775 Online 2320 7876

Research Paper © 2012 IJFANS. All Rights Reserved Journal Volume 13, Iss 4, 2024



Figure-1: Temperature(°C), Turbidity(NTU) and pH of Gundla cheruvu showing Ranges in for the year 2014-2015



Figure-2: Biological Oxygen Demand, Chemical Oxygen Demand and Dissolved Oxygen of Gundla cheruvu showing Ranges expressed in (mg/L) for the year 2014-2015

Table-2:Physico-chemical	parameters	of Gundla	cheruvu	showing	ranges	expressed	in
(mg/L) for the year 2014-20	15						

Range	TDS	TSS	Chlorides	Nitrates	Phosphates	Sulfates	Silicates
Min	122	85.0	77.0	0.42	0.13	19.5	2.1
Max	240	506.0	97.7	0.92	0.90	40.1	5.2

Note: Max=Maximum, Min=Minimum



ISSN PRINT 2319 1775 Online 2320 7876

Research Paper © 2012 IJFANS. All Rights Reserved Journal Volume 13, Iss 4, 2024



Figure-3: Total Dissolved Solids, Total Suspended Solids and Chlorides of Gundla cheruvu showing Ranges expressed in (mg/L) for the year 2014-2015



Figure-4: Nitrates and Phosphates of Gundla cheruvu showing Ranges expressed in (mg/L) for the year 2014-2015



Figure-5: Sulfates and Silicates of Gundla cheruvu showing Ranges expressed in (mg/L) for the year 2014-2015



PHYTOPLANKTON DIVERSITY

Table-3: Distribution of phytoplankton species with respect to class and family in Gundla cheruvu for the year 2014-2015

Gundla Cheruvu (2014-2015)							
S.No	Class	Family	Name of algal forms				
1	Chlorophyceae	Volvocaceae	Pandorina morum (O.F.Müller) Bory				
2		Scenedesmaceae	Scenedesmus quadricauda (Turpin) Brebisson				
3			Scenedesmus acuminatus (Lagerheim) Chodat				
4			Scenedesmus armatus (Chod) G.M.Smith				
5			Scenedesmus bijugatus (Turpin)				
6		Hydrodictyaceae	Pediastrum duplex Meyen				
7			Pediastrum boryanum (Turpin) Meneighini				
8			Pediastrum simplex Meyen				
9			Pediastrum Tetraodon (Corda) A.Braun, nom. illeg.				
10			Pediastrum tetras (Ehrenberg) Ralfs				
11		Desmidaceae	Cosmarium geminatumf. ornatum Behre				
12			Cosmarium subtumidum Nordstedt				
13			Closterium acutum Brébisson				
14			Closterium tumidum f. minus Strøm				
15			Staurastrum javanicumvar. Maximum C. Bernard				
16			Staurastrum manfeldtii Delponte				
17		Selenastraceae	Selenastrum gracile Reinsch				
1	Bacillariophyceae	Naviculaceae	Navicula cuspidata Kutzing				
2			Navicula cincta (Ehr) Kutz				
3		Bacillariaceae	Nitzschia denticula Grunow				
4		Cymbellaceae	Cymbella aspera (Ehrenberg) Cleve				



ISSN PRINT 2319 1775 Online 2320 7876

Research Paper © 2012 IJFANS. All Rights Reserved Journal Volume 13, Iss 4, 2024

5		Achnanthaceae	Achnanthes microcephala(Kützing) Grunow
6			Achnanthes minutissima Kützing
7		Pinnulariaceae	Pinnularia borealis Ehrenberg
8		Fragilariaceae	Gomphonema acceptatum Levkov, Mitic- Kopanja & E. Reichardt
9			Synedra ulna (Nitzsch) Ehrenberg
10			Synedra tabulata (C.Agardh) Kützing
11		Melosiraceae	Melosira granulate (Ehrenberg) Ralfs (Ehrenberg) Ralfs
1	Cyanophyceae	Chroococaceae	Chroococcus minutus var. minimus Keissler
2		Oscillatoriaceae	Oscillatoria formosa var. australica Playfair
3			Oscillatoria curviceps C.Agardh ex Gomont
4			Oscillatoria obtusa gardener
5			Oscillatoria hamelii fremy
6			Anabaena orientalis f. major Laloraya & Mitra
7			Anabaena sphaerica Born et Flax
8		Microcystaceae	Merismopedia glauca (Ehrenberg) Kützing
9		Merismopediaceae	Aphanocapsa annulate G.B.Mc Gregor
1	Euglenophyceae	Euglenaceae	Phacus curvicauda Svirenko
2			Phacus plueronectes (Muell) Dujardin
3			Trachelomonas hispida (Perty) F.Stein

Chlorophyceae

The Chlorophyceae members are categorized as the first high class among phytoplankton group in our present investigation. The results of phytoplankton for Gundla cheruvu, class Chlorophyceae comprises of 5 families, Volvocaceae 1 species Pandorina morum, Scenedesmaceae 4 species Scenedesmus quadricauda, Scenedesmus acuminatus, Scenedesmus armatus, Scenedesmus bijugatus, Hydrodictyaceae 5 species Pediastrum duplex, Pediastrum simplex, Pediastrum boryanum, Pediastrum tetras, Pediastrum tetraodon, Desmidaceae 6 species Closterium acutum, Closterium tumidum, Cosmarium geminatum, Cosmarium subtumidum,



ISSN PRINT 2319 1775 Online 2320 7876

Research Paper © 2012 IJFANS. All Rights Reserved Journal Volume 13, Iss 4, 2024

Staurastrum manfeldtii, Staurastrum javanicum and Selenastraceae 1 species Selenastrum gracile.

Bacillariophyceae

The results of phytoplankton for Gundla cheruvu, class Bacillariophyceae comprises of 7 families, Naviculaceae 2 species Navicula cuspidate, Navicula cincta, Bacillariaceae 1 species Nitzschia denticule, Cymbellaceae 1 species Cymbella aspera, Achnanthaceae 2 species Achnanthes microcephala, Achnanthes minutissima, Pinnulariaceae 1 species Pinnularia borealis, Fragilariaceae 3 species Synedra tabulate, Synedra ulna, Gomphonema acceptatum and Melosiraceae 1 species Melosira granulate.

Cyanophyceae

The results of phytoplankton for Gundla cheruvu, class Cyanophyceae comprises of 4 families, Chroococaceae 1 species Chroococcus minutus, Oscillatoriaceae 6 species Oscillatoria formosa, Oscillatoria curviceps, Oscillatoria obtusa, Oscillatoria hamelii, Anabaena orientalis, Anabaena sphaerica,, Microcystaceae 1 species Merismopedia glauca and Merismopediaceae 1 species Aphanocapsa annulate.

Euglenophyceae

The results of phytoplankton for Gundla cheruvu, class Euglenophyceae comprises of 1 family, Phacus curvicauda, Phacus plueronectes and Trachelomonas hispida.

Table-4:	Total	values	and	Percentage(%) 0	f phytoplanton	in	Gundla	cheruvu	from	2014-
2015											

Phytoplanton	2014-2015	Percentage%
Chlorophyceae	3029	47
Bacillariophyceae	1697	26
Cyanophyceae	1327	21
Euglenophyceae	374	6



IJFANS INTERNATIONAL JOURNAL OF FOOD AND NUTRITIONAL SCIENCES ISSN PRINT 2319 1775 Online 2320 7876 Research Paper © 2012 IJFANS. All Rights Reserved Journal Volume 13, Jss 4, 2024



Figure:6 Total values and Percentage (%) of phytoplanton in Gundla cheruvu from 2014-2015

The members of Chlorophyceae class occupied first position in order of dominance in Gundla cheruvu for November 2014 to October 2015 with 3029. The percentage recorded as 47%. Bacillariophyceae recorded 1697 with the percentage of 26% followed by Cyanophyceae 1327 with 21% and the least Euglenophyceae 374 with 6%, as shown in Table-5 and Figure-4, similar to Nygaard (1949), Abdar (2013), Round (1957), Ghosh et al., (2012), Tiwari & Chauhan (2006), Ali, Abd el-Salam (1999) and Lurling et al., (2013).

CONCLUSION

The results indicated a trend towards increasing eutrophication, suggesting that Gundla cheruvu is moderately polluted. The growth of phytoplankton was facilitated by the abundance of phosphates and nitrogen. The distribution of phytoplankton varied with seasons: Chlorophyceae dominated during the summer and pre-monsoon, Bacillariophyceae were the second most dominant species in the pre-monsoon, Cyanophyceae were prevalent in the summer, and Euglenophyceae dominated in the post-monsoon. The eutrophic condition of Gundla cheruvu was reflected in various physico-chemical parameters, including water temperature, turbidity, pH, biological oxygen demand (BOD), chemical oxygen demand (COD), dissolved oxygen (DO), total dissolved solids (TDS), total suspended solids (TSS), and concentrations of chloride, nitrate, phosphate, sulphates, and silicates. To maintain the health of these water bodies, appropriate maintenance is essential. Environmental education and proper sanitation practices are crucial to keeping these waters safe and clean. Simple measures, such as redirecting sewage and preventing nutrient leaching from the catchment area through planting vegetation, can significantly contribute to a clean and sustainable environment.



ISSN PRINT 2319 1775 Online 2320 7876

Research Paper © 2012 IJFANS. All Rights Reserved Journal Volume 13, Iss 4, 2024

AKNOWLEDGEMENT

The authors are grateful to Head department of Botany and Osmania university for valuable suggestions and constant encouragement.

REFERENCES

1. Abdar, M.R. (2013). Physico-chemical characteristics and phytoplankton of Morna lake, Shirala (M.S.) India, Biolife, Vol. 1 (2): 1-7.

2. Abd-Ellatif, M., Howayda, H., El-Hady, A., Toufeek, M.E.F. and Varbiro, G. 2016. Phytoplankton structure, biochemical, stoichiometry and elemental composition in Lake Nasser, Egypt. Int. J. Appl. Environ. Sci. 11: 211-228.

3. Agarwal, S.C. (1999) . Limnology, A.P.H, Publishing Corporation, New Delhi. pp 3-9.

4. Airill, L.M., Querijero, B.L. and Ching, J.A. 2016. Phytoplankton community in aquaculture and non-aquaculture sites of Taal lake, Batangas, Philippines. J. Exper. Biol. Agri. Sci. 4: 66-73.

5. Ali GH, Abd el-Salam NF (1999) Factors controlling bioindicator for industrial pollution detection. Biomed Environ Sci 12:194-200.

6. Altaf H. Ganai and Saltanat Parveen(2014), "Effect of physicochemical conditions on the structure and composition of the phytoplankton community in Wular Lake at Lankrishipora, Kashmir", International Journal of Biodiversity Conservation, 6 (1), 71-84.

7. Amin Hossaini Motlagh, Navatha K and Manikya Reddy P.,(2013)."Ecological Studies of Mir Alam Lake with Reference to Water Quality", Nature Environment and Pollution Technology;12, 355-358.

8. Arruda, J. A., Marzolf, G. R., & Faulk, R. T. (1983). The role of suspended sediments in the nutrition of zooplankton in turbid reservoirs. Ecology, 64(5), 1225-1235.

9. Arumugam, V., Sivakami, R. and Premkishore, G. 2015. Biodiversity of phytoplankton in a tropical lake of South India. Int. J. Curr. Microbiol. Appl. Sci. 4: 362-376.

10. B.K. Dwivedi and G.C. Pandey. 2002. Physico-Chemical factors and algal diversity of two ponds, (Girija Kund and Maqubara pond), Faizabad. Poll. Res., 21, 361-370.

11. Berry, W., Rubinstein, N., Melzian, B., & Hill, B. (2003). The biological effects of suspended and bedded sediment (SABS) in aquatic systems: a review. United States Environmental Protection Agency, Duluth, 32(1), 54-55.

12. Bhalla, Resham, V.S. Lomte and M.B. Mule, 2006. Physico-Chemical Assessment of water in relation to the primary production of planktons of Godavari river at Nasik. Bull.Env.Sci. vol.xxiv (2nd issue) pp. 165-169.9).

13. Bowling, L (2009). Freshwater phytoplankton: diversity and biology.

14. Boyd, P. W., Rynearson, T. A., Armstrong, E. A., Fu, F., Hayashi, K., Hu, Z., et al. (2013). Marine phytoplankton temperature versus growth responses from polar to tropical waters– Outcome of a scientific community-wide study. PLoS ONE 8:e63091.

15. Caux, P. Y., Moore, D. R. J., & MacDonald, D. (1997). Ambient water quality guidelines (criteria) for turbidity, suspended and benthic sediments: technical appendix. Prepared for BC Ministry of Environment, Land and Parks (now called Ministry of Water, Land and Air Protection).



ISSN PRINT 2319 1775 Online 2320 7876

Research Paper © 2012 IJFANS. All Rights Reserved Journal Volume 13, Iss 4, 2024

16. Chapman, P. M., Hayward, A., & Faithful, J. (2017). Total suspended solids effects on freshwater lake biota other than fish. Bulletin of environmental contamination and toxicology, 99, 423-427.

17. Chatterjee, A.K.K.1992.Water quality of Nandankanan. Indian J. Environ.Hlt., 34(4): 329-333.

18. Chinnaiah B Ramesh Babu M and B.Digamber Rao.2011. Phycoplankton diversity and population dynamics of Ramappa Lake,(A.P) India.Ad.Plant.Sci.24(II):527-529.

19. Das SM, Pandey J (1978). Lake pollution as evidenced by physical, biological and chemical parameters in the Nainital lake, U.P. In: Proceedings of the national seminar on DST, New Delhi, pp 502–512.

20. EPA (2001). United States environmental protection agency. Quality Assurance Guidance Document-Model Quality Assurance Project Plan for the PM Ambient Air, 2, 12.

21. Garg, R. K., Saksena, D. N. and Rao, R. J. (2006). Assessment of physic-chemical water quality of Harsi Reservoir, district Gwalior, Madhya Pradesh. Jour. Ecophysiol. Occupat. Heal. 6:33-40.

22. Ghosh, S., Barinova, S. and Keshri, J.P. 2012. Diversity and seasonal variation of phytoplankton community in the Santragachi Lake, West Bengal, India. Q Sci. Connect. 3: 1-13.

23. Hascoet MC, Jarret M & Ducauze CJ (1986). Evolution de la matiére organique lors de la filtration sur C.A.G. Incidence de l'ozonation. Sci. Tech. Eau. 5: 197-213.

24. Ishaq, F., Khan, A., (2013): indicator for water quality criteria of river Yamuna in Doon Valley, Uttarakhand, India. World J. Fish Mar. Sci.5, 322–334.

25. Jones, W. B., Romain, K., Erlandson, R. A., Burt, M. E., & Lewis Jr, J. L. (1993). Thoracotomy in the management of gestational choriocarcinoma. A clinicopathologic study. Cancer, 72(7), 2175-2181.

26. Kannel, P.R., Lee, S., Lee, Y.S., Kanel, S.R. and Pelletier, G.J. (2007) Application of Automated QUAL2Kw for Water Quality Modeling and Management in the Bagmati River, Nepal. Ecological Modelling, 202, 503-517.

27. Khound Nayan J Phukon P, Bhattacharyya KG (2012). A comparative study of ground water and surface water quality in the Jia-Bharali river basin, Assam, India with reference to Physico-chemical characteristics. Int J Appl Sci Eng Res 1(3):512–5.

28. Krishnamoorthi, A., Senthil Elango, P. & Selvakumar, S. (2011). Investigation of water quality parameters for aquaculture A case study of veeranam in Cuddalore district, Tamil Nadu. International Journal of Current Research, 3(3), 13-17.

29. Kumar. M. P., Prabhahar. C., (2012): Physicochemical parameters of river water a review. Int. J. Pharm. Biol. Arch.3.1304: 1312.

30. Kumbhar, A. C., & Bade, D. K. B. (2009). SeasonalVariation In Physico-Chemical Parameters Of Ujani Reservoir Of Madha Tahashil, Dist, Solapur. Shodh, Samoksha Aur Mulankan (International Research Journal) Vol. II,(7), 150-152.

31. Lalparmawii, S. (2007). Analysis of water quality and Biomonitoring of Tuirial River in vicinity of the Hydel Project in Mizoram. Ph.D Thesis. Mizoram University, Aizawl.



ISSN PRINT 2319 1775 Online 2320 7876

Research Paper © 2012 IJFANS. All Rights Reserved Journal Volume 13, Iss 4, 2024

32. Lloyd, D. S. (1985). Turbidity in freshwater habitats of Alaska: a review of published and unpublished literature relevant to the use of turbidity as a water quality standard. Alaska Department of Fish and Game, Habitat Division.

33. Lurling, M., Eshetu, F., Faassen, E.J., Kosten, S. and Huszar, V.L.M. 2013. Comparison of cyanobacterial and green algal growth rates at different temperatures. Freshw. Biol. 58: 552-559.

34. Mithani, I., Dahegaonkar, N. R., Shinde, J. S. and Tummawar, S. D. (2012). Preliminary studies on physico-chemical parameters of river Wardha, District Chandrapur, Maharashtra. Jour. Res. Ecol. 1: 14-18.

35. Mittal, S. and Sengar, R. M. S., 1989. Toxic effect of sulphate and its uptake in algae. Natl Acad Sci Lett, 12:17-19.

36. Nygaard, G., 1949. Hydrobiological studies on some Danish ponds and lakes 11. The quotient hypothesis and some new or little known phytoplankton organisms. Dat. Kurgl. Danske. Vial. Sel. Biol. Skr., 7: 1 -293.

37. Panday S and Tripati A.K. (1984). Algal pollutants of Unnao ponds II. Qualitative, Quantitative and periodical occurrence of Cyanophyceae. J. Pl. Nature. 1(1):83-86.

38. Rai, L.C. 1978. Ecological studies of algal communities of the ganges river at Varanasi. India. J.Ecol.5:1-6.

39. Rajakumar R and Ramanibai R (1994), Observations of species (plankton) environmental relationship in urban aquatic ecosystem. J. Environ. Bio. 15(3) :177-183.

40. Rajashekar, M., Vijaykumar, K. and Haliked, N. S. (2010). Seasonal variation in physicochemical parameters of Hirahalla reservoir, Koppa District Karnataka. Internat. Jour. Syst. Bio. 2(2): 16-20. 48.

41. Ross, D. A. (1969). Temperature structure of the Red Sea brines. Hot Brines and Recent Heavy Metal Deposits in the Red Sea: A Geochemical and Geophysical Account, 148-152.

42. Round, F.E., 1957. Studies on bottom living algae in some lakes of English Lake District. Part I. Some chemical features of sediments related to algal productivities. J. Ecol. 45: 133 -48.

43. Saifullah, A. S. M., Hena, M. K. A., Idris, M. H., Halimah, A. R. and Johan, I. (2014). Diversity of phytoplankton from mangrove Estuaries of Sarawak, Malaysia. World Appl. Sci. J., 31: 915–924.

44. Sharma, P., Sood, A., Sharma, S., Bisht, S., Kumar, V., Pandey, et al. 2010. Bacterial indicators of faecal pollution and physiochemical assessment of important North Indian lakes. RMZ – Mat. Geoenvir. 57: 25-40.

45. Shashikant and Anil. K. Raina. (1990). Limnological studies in Jammu.11.Physico-chemical parameters. J. Environ. Biol.,11(2):137.

46. Shuhaimi-Othman.M and A.K. Ahmad 2010. Heavy Metal Concentrations in Sediments and Fishes from Lake Chini, Pahang, Malaysia. Journal of Biological Sciences, 10: 93-100.

47. Singh, M.R. and Gupta, A. (2010). Seasonal variation in certain physicochemical parameters of Imphal, Iril and Thoubal rivers from Manipur river system, India. Ecology, Environment and Conservation, 16(2): 197-207.

48. Sinha SN, Paul D and Biswas K (2016): Effects of Moringa oleifera Lam. and Azadirachta indica A. Juss. leaf extract in treatment of tannery effluent. Our Nature; 14(1): 47-53.



ISSN PRINT 2319 1775 Online 2320 7876

Research Paper © 2012 IJFANS. All Rights Reserved Journal Volume 13, Iss 4, 2024

49. Solanki D.L., Vasudha L., Anuradha, D.L. and Sabita Raja S. (2015). Water quality assessment of Nallacheruvu, Hyderabad, Telangana, India. Journal of Bio Research 1(1):1-12.

50. Subba Rao M. V., Rao B. M. G., Rao B.R. and Nanda N. K (1981). Hydrological studies of the brackish water Chilika lagoon, Orissa. Journal of Environmental Biology 2:59-62.

51. Swarnalatha, N. and A. Narsing Rao, 1998: Ecological studies of Banjara lake with reference to water pollution. Journal of Environmental Biology 19: 179-186.

52. Thomas, G. A., Phillips, T. G., Rice, T. M., & Hensel, J. C. (1973). Temperature-dependent luminescence from the electron-hole liquid in Ge. Physical Review Letters, 31(6), 386.

53. Tiwari, A. & Chauhan, S.V.S. (2006). Seasonal phytoplanktonic diversity of Kitham Lake, Agra, Journal of Environmental Biology, 27: 35-38.

54. Trivedy, R. K., Khatavkar, S. D., Kulkarni, A. Y. and Shirotri, A. C. (1990). Ecology and pollution of river Krishna in Maharastra. I: General features of the river and pollution inventory. In: River pollution in India. Trivedy, R. K. (Ed.). Ashish Publishing House, New Delhi. 71-133.

55. Wani, S. H., Singh, N. B., Devi, T. R., Haribhushan, A., Jeberson, S. M., Malik, C. P., et al. (2013). "Engineering abiotic stress tolerance in plants: extricating regulatory gene complex," in Conventional and Non-Conventional Interventions in Crop Improvement, eds C. P. Malik, G. S. Sanghera, and S. H. Wani (New Delhi: CABI), 1–19.

56. Warwick, R. M. (1983). The implications of resource partitioning for the structure of a sandbeach meiofauna community. In Sandy Beaches as Ecosystems: Based on the Proceedings of the First International Symposium on Sandy Beaches, held in Port Elizabeth, South Africa, 17–21 January 1983 (pp. 753-754). Dordrecht: Springer Netherlands.

57. Young A, Robert, Donald J. P. Swift, Thomas L, Clarky, George R, Harvey and Peter R Betzer (1985). Dispersal pathways for particle associated pollutants. Science. 8. 229, 4712. 431-435.

