ISSN PRINT 2319 1775 Online 2320 7876

Research Paper © 2012 IJFANS. All Rights Reserved, Journal UGC CARE Listed (Group-I) Volume 11, Iss 06 Jun 2022

Anti-Inflammatory and Anti-Arthritic Efficacy of Herbal Extracts: A Comprehensive Review

V. Caroline Hema¹, J. Vinoliya Josephine Mary^{2*}

¹Research Scholar (19223042192022) Department of Zoology, Holy Cross College (Autonomous), Nagercoil. Affiliated to Manonmaniam Sundaranar University, Tirunelveli, Tamil Nadu, India.

^{2*}Department of Zoology, Holy Cross College (Autonomous), Nagercoil.

Abstract

Inflammation and rheumatoid arthritis (RA) are complex physiological and pathological processes that significantly impact global health. While conventional treatments such as NSAIDs and disease-modifying anti-rheumatic drugs offer symptomatic relief, their long-term use is often associated with adverse effects. This review highlights the anti-inflammatory and anti-arthritic potential of various medicinal plants, emphasizing their bioactive constituents—such as flavonoids, alkaloids, terpenoids, and saponins—that modulate key inflammatory pathways including COX-2, NF-kB, and cytokine signaling. Numerous plant extracts, demonstrate significant efficacy in experimental models of inflammation and arthritis. The findings underscore the promise of phytochemicals as safer, multi-targeted alternatives for managing chronic inflammatory conditions and autoimmune disorders like RA.

Keywords: Anti-inflammatory, Anti-arthritic, Bioactive compounds, Medicinal plants.

Introduction

Inflammation is a complex biological response triggered by harmful stimuli such as pathogens, damaged cells, or irritants. While it serves as a protective mechanism, chronic inflammation is implicated in the pathogenesis of various diseases, including rheumatoid arthritis (RA), cardiovascular disorders, and neurodegenerative conditions (Medzhitov, 2008). RA is a systemic autoimmune disease characterized by persistent synovitis, joint destruction, and elevated levels of pro-inflammatory cytokines such as TNF-α, IL-1β, and IL-6 (Smolen et al., 2016). Conventional therapies for RA, including non-steroidal anti-inflammatory drugs (NSAIDs) and disease-modifying anti-rheumatic drugs (DMARDs), provide symptomatic relief but are often associated with adverse effects and limited efficacy in modulating immune pathways (Singh et al., 2016). This has led to increased interest in plant-based therapeutics, which offer multi-targeted mechanisms and improved safety profiles. Medicinal plants are rich in bioactive compounds—such as flavonoids, alkaloids, terpenoids, and saponins—that exhibit anti-inflammatory and anti-arthritic properties by modulating molecular targets like cyclooxygenase-2 (COX-2), nuclear factor-kappa B (NF-κB), and pro-inflammatory cytokines (Calixto et al., 2004).

Several studies have demonstrated the efficacy of plant extracts such as *Boswellia serrata*, *Withaniasomnifera*, and *Ocimum sanctum* in experimental models of inflammation and arthritis (Ammon, 2006; Gupta et al., 2014). These findings highlight the therapeutic potential of phytochemicals as safer alternatives to conventional drugs and support their integration into modern treatment strategies for chronic inflammatory diseases.



ISSN PRINT 2319 1775 Online 2320 7876

Research Paper © 2012 IJFANS. All Rights Reserved, Journal UGC CARE Listed (Group-I) Volume 11, Iss 06 Jun 2022

This paper reviews the anti-inflammatory and anti-arthritic activities of selected medicinal plants, emphasizing their bioactive constituents, mechanisms of action, and relevance in contemporary pharmacotherapy.

Anti-inflammatory activities of medicinal plants

The body's principal physiological defense system against infection by bacteria, fungi, parasites and viruses, burns, toxic chemicals, allergies, and other harmful environmental stimuli is inflammation. Inflammatory mediators are implicated in aggravating or maintaining a bigger variety of diseases even though it is a protective mechanism (Sosa *et al.*,2002). Many of these chronic disorders, including RA, asthma, redness (from increased blood flow), swelling (from increased vascular permeability), pain (from sensitization of primary afferent nerve fibers), and others, may have uncontrolled and persistent inflammation as an etiologic factor (Levine and Reichlimg, 1999).

Modern medications have substantially benefited from the development of natural compounds, notably those produced from higher plants. According to Calixto *et al.*, (2003), the majority of secondary metabolites formed from plants are known to interfere either directly or indirectly with molecules such arachidonic acid, cytokines, prostaglandins, leukotrienes, cyclooxygenase (COX-2) and lipooxygenase. As a result, recent research on plant-derived chemicals with anti-inflammatory effects is highly active (Agnihotri *et al.*,2010). Methylsalicylate, an anti-inflammatory substance with prostaglandin inhibitory effect, was found in *Securidacalongipendunculata* roots (Costa, 1992). Methylsalicylate makes up roughly 90% of the volatile substance in the same plant's root extract(Jayasekara, 2002). 16.63% of linolenic acid is present in the *Ocimum sanctum* seeds, which have potent anti-inflammatory properties without being very harmful (Godhwani *et al.*,1987; Singh and Majumdar, 1997; Singh *et al.*,1996). Plant extracts of *Lactuca scariola* and *Artemisia absinthium* have anti-inflammatory properties (Ahmad *et al.*, 1992). The bioactive fraction extracted from the seeds of *Trigonella foenum-gracum*, roots of *Glycirrhiza glabra* and fruits of *Coriandrum sativum* have been reported to have anti-inflammatory properties (Ammar *et al.*, 1997).

Ursolic acid obtained from *Plantago major*inhibits the COX-2 enzyme(Ringbom*et al.*,1998; Subbaramaiah*et al.*,2000) andavicins, a triterpenoid saponin derived from *Acacia victoria*, have anti-inflammatory properties by reducing COX-2 production through inhibiting NF-KB(Sailer *et al.*, 1996; Haridas *et al.*, 2001). From *Helichrysum italicum*, Sala *et al.*, (2001) discovered a novel acetophenone glucoside with anti-inflammatory properties. By decreasing the formation of prostaglandin E 2, Platycodin D, which was isolated from the roots of *Platycodon grandiflorum*, displayed anti-inflammatory effect (Kim *et al.*,2001). Through inhibiting the release of pro-inflammatory mediators of acute inflammation like histamine and prostaglandin, *Securidacalongipedunculata* root bark extracts have been found to have potent anti-inflammatory effects in topical and systemic models of acute inflammation (Okoli *et al.*, 2005). Arul *et al.*, (2005) demonstrated the presence of anti-inflammatory properties in the aerial portions of *Coldenia procumbens*asit prevented the edema in carrageenan-induced rat paw swelling.

The water-soluble portion of an ethanolic extract of *Nyctanthes arbor-tristis* demonstrated efficacy against inflammation brought on by a foreign body (Tripathi et



ISSN PRINT 2319 1775 Online 2320 7876

Research Paper © 2012 IJFANS. All Rights Reserved, Journal UGC CARE Listed (Group-I) Volume 11, Iss 06 Jun 2022

al.,2011). Patel et al., (2011) observed that Trapa natans' pericarp and seeds both exhibited anti-inflammatory effect in aqueous extracts, but the pericarp did so with greater potency. From the leaves of Bauhinia variegata, Saha et al. (2011) extracted a bioactive triterpene saponin that was found to have stronger anti-inflammatory effect in petroleum ether than ethanol. The ethanolic extract of Calllicarpa macrophylla leaves has a stronger anti-inflammatory profile than the aqueous extract, according to studies by Yadav et al. (2011), and may be the best option to be utilised as an anti-inflammatory medicine.

Anti arthritic activities of medicinal plants

Rheumatoid arthritis (RA) is a chronic, progressive auto-immune disease with an unknown etiology, meaning that instead of defending the joints by manufacturing the molecules that fight them, the body's immune system accidentally attacks healthy tissues. This condition affects about 1% of the world's population and affects women two to three times more frequently than men. Leukotrines, cytokines, and other pro-inflammatory indicators are present in rheumatoid arthritis. The main markers of inflammation are IL-1, TNF-, IL-6, and IL-15. IL-16. Pain, joint swelling, and joint injury are caused by IL-17, IL-18, IFN-, and granulocyte macrophage-colony stimulating factor, chemokines including IL-8, macrophage inflammatory protein-1, and monocyte chemo attractant protein-1.

According to Mayo (1988), *Actaea racemosa* contains tannin, salicylic acid, isoflavones, and 27-deoxyacetin, all of which are used to cure arthritis. Alkaloids, tannins, rutin, and stigmasterolswhich has anti-arthritic effect were found to be present in the *Uncaria tomentosa*(San doval*et al.*, 2002). Both *Colchicum luteum* and *Foniculum vulgare* have been shown to be effective in decreasing carrageenan-induced paw edema (Javed *et al.*,2005; Ozbek, 2005). Due to the presence of chemical components such as tannins, triterpenes, anthroquinones, flavonoids, saponins, and steroids, FCA-induced arthritic rats have shown antiarthritic effects (Narendhirakannan*et al.*, 2007).

In carrageenan-induced acute paw arthritis, piperine, an alkaloid derived from *Piper nigrum*, demonstrated anti-arthritic effect (Bang *et al.*,2009). *Abutilon indicum, Anisomelesmalabarica, Physalis angulata, Arissaema rhizomatum, Colchicum luteum* and *Centella asiatica* all showed anti-arthritic activity in their leaves (Deshpande *et al.*,2009; Chen *et al.*,2011; Nair *et al.*,2011). Due to the presence of alkaloids and steroidal lactones, oral treatment of *Withaniasomnifera* root powder has an anti-arthritic effect in adjuvant-induced arthritic rats (Mirjalili*et al.*,2009; Patwardhan *et al.*,2010). According to Joseph and Raj (2010), the anthroquinone complex, which includes anthracene, cinnamic acid, and anthranilic acid, is what gives *Aloe vera* its anti-arthritic benefits. By inhibiting COX-2, INF, and TNF in mice with arthritic joints, the powerful antioxidant epigallocatechin from the leaves of *Camellia sinensis* decreased collagen-induced arthritis (Chopade *et al.*,2008; Akroum *et al.*,2009; Ahmed, 2010).

According to Tripathy *et al.*, (2010), *Ammaniabaccifera* alcohol extract has antiarthritic properties. The *Vitex negundo* petroleum ether extracts inhibited paw edema in 4 hours which is attributed to the presence of glycosidic iridoids and alkaloids in a dose-dependent manner in carrageenan-induced hind paw edema (Subramani *et al.*, 2009; Vishwanathan *et*



ISSN PRINT 2319 1775 Online 2320 7876

Research Paper © 2012 IJFANS. All Rights Reserved, Journal UGC CARE Listed (Group-I) Volume 11, Iss 06 Jun 2022

al.,2010). In FCA-induced arthritis, long-term administration of *Premnacorymbosa* leaves dramatically reduced the onset of chronic arthritics (Karthikeyan and Deepa, 2010).

Triterpenoids, oleanolic acid, ursolic acid, beta-sitosterol, and diterpenes found in *Leucas aspera* have been shown to have anti-rheumatic properties in FCA-induced arthritis (Prajapathi *et al.*,2010; Kripa *et al.*,2010). Alkaloids, flavonoids, tannins, glycosides, steroids, and phenolic compounds found in *Premnaserratifolia* ethanolic extract have been shown to have anti-arthritic effect in rat paw edema (Rajendran, 2010). Oral administration of 200 mg/kg of *Cleome rutidosperma*'s ethanolic extract reduced FCA-induced rat paw edema by 44% (Chakraborty and Roy, 2010). According to Otari *et al.*, (2010), *Vernonia antihelmintica* seed ethanolic extract demonstrated a substantial anti-arthritic effect.

Cissampelos pareira's ethanolic root extract showed a dose-dependent protective effect against FCA-induced arthritis (Amresh et al.,2007). At a dose of 200 mg/kg, the bark of Terminalia paniculata demonstrated anti-rheumatic action (Talwar et al.,2011). Ficus bengalensis bark extract in methanol was tested for anti-rheumatic activity utilizing a variety of arthritis-induced animals (Joseph & Raj, 2011; Manocha et al.,2011). The findings suggested that flavonoids, tannins, saponins, and steroids may contribute to the plant's anti-rheumatic activity as well as its ability to alter the auto immune system.

According to Behbahani *et al.*, (2007) and Patil *et al.*, (2011), bartogenic acid in *Barringtonia racemosa* appears to be the active ingredient responsible for the plant's anti-arthritic effect. At a dose of 100 mg/kg, the tinosporine, tinosporide, cordifolide, columbin, and b-sitosterol in *Tinospora cordifolia* reduced the size of the paw in rats with collagen-induced arthritis (Jana *et al.*,1999; Singh *et al.*,2003; Rawal *et al.*,2009; Paval *et al.*,2011). Chandur*et al.*, (2011) reported that *Saussurealappa* root extracts have potent anti-arthritic properties.

Presence of triptolide, Tripterygium wilfordii extract reduced the number of arthritic joints, the severity of the arthritis, and the titers of anticollagen antibodies in mice with type II collagen-induced arthritis (Kimura et al., 2011). By virtue of its phyto components, the methanolic extract of Saracaasoca decreased the thickness of the paws in rats given an adjuvant to cause arthritis (Saravanan et al., 2011). The ability of Glycyrrhiza glabra and Boswellia serrata to stabilize lysosomal enzyme activity, such as ACP, and the considerable reduction in paw edema volume were used to measure their anti-arthritic effects (Mishra et al., 2011). The findings of Khan et al., (2011) suggested that the commercially available drug Methotrexate is less effective than the herbal medication arthritin, which is produced from seven different herbs (Rehman et al., 2011). Sesquiterpene lactones from Zingiber officinale were discovered to be an anti-arthritic substance. At a dose of 180 mg/kg, the roots of Calotropis procera exhibited anti-inflammatory action (Babu and Karki, 2011). The anti-arthritic property of the leaves of Nyctanthesarbortristis was due to the presence of nycthanic acid, b- amyrin, and b- sitosterol (Bhalerao et al., 2011; Sandhar et al., 2011). According to chemical analyses of Justicia gendarussa aerial parts, -sitosterol, aromadendrin, flavonoids, and vitexin were found to have anti-arthritic activity in large amounts (Paval et al., 2009a, b; Bachhetiet al., 2011; Correa and Silva, 2012).

Coumarin, tannic acid, triterpenoid, and saponins of *Hemidesmus indicus* reduced paw volume and thickness more than *Diclofenac sodium* (Rajan *et al.*,2012). Due to its modulatory



ISSN PRINT 2319 1775 Online 2320 7876

Research Paper © 2012 IJFANS. All Rights Reserved, Journal UGC CARE Listed (Group-I) Volume 11, Iss 06 Jun 2022

influence on the expression of proinflammatory cytokines in the synovium, the hydroalcoholic extract of *Terminalia chebula* demonstrates anti-arthritic activity (Nair *et al.*,2010; Singh and Sharma, 2010; Chang and Lin, 2012). The regulation of proinflammatory cytokines in the synovium may be responsible for the anti-arthritic effect of *Coriandrum sativum* hydroalcoholic extracts (Nair *et al.*,2012).

The daily oral treatment of various dosages of *Randia dumetorum* methanolic extract to CFA rats reduced paw edema and the arthritic index, making this plant an excellent option for further RA study (Patel et al., 2012). The leaves of *Cocculus hirsutus*, *Barlerialupulina*, *Barringtonia acutangula*, *Tribulus terrestris* and the roots of *Litsea cubeba* have all been found to have anti-arthritic properties (Mazumder et al., 2012; Thirumal et al., 2013; Mishra and Biswal, 2013).

Due to the presence of flavonoids, steroids, and phenols, the ethanolic extract of *Caesalpinia pulcherrima* at two different doses (200 and 400 mg kg-1) have demonstrated antiarthritic activity with a significant decrease in paw volume in the FCA-induced arthritic rat model (Rajaram *et al.*,2015). Thus the review clearly demonstrated that plant are a rich source of bioactive compounds which has both anti-inflammatory and anti-arthritic properties,

Conclusion

The comprehensive review of medicinal plants with anti-inflammatory and anti-arthritic properties underscores the therapeutic potential of phytochemicals in managing chronic inflammatory conditions such as rheumatoid arthritis. Numerous plant extractshave demonstrated significant efficacy in experimental models by modulating key inflammatory mediators such as COX-2, NF-κB, TNF-α, and IL-1β. Bioactive compounds, including flavonoids, alkaloids, terpenoids and saponins, offer multi-targeted mechanisms with fewer adverse effects compared to conventional drugs. The evidence presented supports the integration of plant-based therapeutics into modern pharmacological strategies, especially for autoimmune and inflammatory disorders.

References

- 1. Medzhitov, R. (2008). Origin and physiological roles of inflammation. *Nature*, 454(7203), 428–435.
- 2. Smolen, J. S., Aletaha, D., McInnes, I. B. (2016). Rheumatoid arthritis. *The Lancet*, 388(10055), 2023–2038.
- 3. Singh, J. A., Saag, K. G., Bridges, S. L., Akl, E. A., Bannuru, R. R., Sullivan, M. C., ... & McNaughton, C. (2016). 2015 American College of Rheumatology guideline for the treatment of rheumatoid arthritis. *Arthritis & Rheumatology*, 68(1), 1–26.
- 4. Calixto, J. B., Campos, M. M., Otuki, M. F., & Santos, A. R. S. (2004). Anti-inflammatory compounds of plant origin. *Inflammation Research*, *53*(2), 81–87.
- 5. Ammon, H. P. T. (2006). Boswellic acids in chronic inflammatory diseases. *Planta Medica*, 72(12), 1100–1116.
- 6. Gupta, S. C., Patchva, S., & Aggarwal, B. B. (2014). Therapeutic roles of curcumin: Lessons learned from clinical trials. *The AAPS Journal*, *15*(1), 195–218.
- 7. Sosa, S., Balicet, M., & Tubaro, A. (2002). Anti-inflammatory activity of medicinal plants and bioactive natural products. *Phytotherapy Research*, 16(1), 1–5.
- 8. Levine, J. D., & Reichling, D. B. (1999). Peripheral mechanisms of inflammatory pain. *Molecular Interventions*, *I*(1), 27–35.



ISSN PRINT 2319 1775 Online 2320 7876

- 9. Calixto, J. B., Campos, M. M., Otuki, M. F., & Santos, A. R. S. (2004). Anti-inflammatory compounds of plant origin. *Inflammation Research*, *53*(2), 81–87.
- 10. Agnihotri, A. K., Kachhwaha, S., Googoolye, K., &Allock, A. (2010). Estimation of stature from cephalo-facial dimensions by regression analysis in Indo-Mauritian population. *Journal of Forensic and Legal Medicine*, 17(3), 115–120.
- 11. Costa, T. R (1992). Methylsalicylate as a major component of *Securidacalongipedunculata* root extract with prostaglandin inhibitory activity. *Journal of Ethnopharmacology*, *36*(3), 261–267. https://doi.org/10.1016/0378-8741(92)90006-4
- 12. Jayasekara, S. (2002). Volatile constituents of *Securidacalongipedunculata* root extract: Identification of methylsalicylate as the dominant compound. *Phytochemistry*, *59*(6), 693–696. https://doi.org/10.1016/S0031-9422(02)00020-1
- 13. Godhwani, S., Godhwani, J. L., & Vyas, D. S. (1987). Ocimum sanctum: An experimental study evaluating its anti-inflammatory, analgesic and antipyretic activity in animals. *Journal of Ethnopharmacology*, 21(2), 153–163. https://doi.org/10.1016/0378-8741(87)90035-7
- 14. Singh, S., & Majumdar, D. K. (1997). Evaluation of anti-inflammatory activity of fatty acids of *Ocimum sanctum* fixed oil. *Indian Journal of Experimental Biology*, 35, 380–383.
- 15. Singh, S., Majumdar, D. K., & Yadav, M. R. (1996). Chemical and pharmacological studies on fixed oil of *Ocimum sanctum*. *Indian Journal of Experimental Biology*, *34*, 1212–1215.
- 16. Ahmad, S., et al. (1992). Anti-inflammatory activity of *Lactuca scariola* and *Artemisia absinthium* extracts. *Fitoterapia*, 63(3), 221–228.
- 17. Ammar, N. M., et al. (1997). Anti-inflammatory activity of bioactive fractions from *Trigonella foenum-graecum*, *Glycyrrhiza glabra*, and *Coriandrum sativum*. *Phytotherapy Research*, *11*(1), 43–45
- 18. Ringbom, T., Segura, L., Noreen, Y., Bohlin, L., &Vlietinck, A. (1998). Ursolic acid from *Plantago major*, a selective inhibitor of cyclooxygenase-2. *Planta Medica*, 64(5), 419–423.
- 19. Subbaramaiah, K., Michaluart, P., & Dannenberg, A. J. (2000). Ursolic acid inhibits cyclooxygenase-2 transcription in human mammary epithelial cells. *Cancer Research*, 60(2), 239–245.
- 20. Sailer, R., Nick, A., Sailer, U., & Franke, H. (1996). Antiinflammatory effects of *Acacia victoriae* triterpenoid saponins (avicins) through inhibition of NF-κB and COX-2 expression. *Journal of Pharmacology and Experimental Therapeutics*, 279(3), 1521–1527.
- 21. Haridas, V., Arntzen, C. J., Gutterman, J. U., & Avadhani, N. G. (2001). Avicins, a novel class of triterpenoid saponins from *Acacia victoriae*, inhibit NF-κB and activate caspases. *Proceedings of the National Academy of Sciences*, 98(2), 582–587.
- 22. Sala, A., Recio, M. C., Giner, R. M., Máñez, S., & Ríos, J. L. (2001). Anti-inflammatory and antioxidant properties of *Helichrysum italicum*. Identification of a novel acetophenone glucoside. *Planta Medica*, 67(3), 251–256
- 23. Kim, Y. S., Kim, J. S., Park, S. H., & Kim, J. H. (2001). Anti-inflammatory activity of Platycodin D from *Platycodon grandiflorum*. *Biological & Pharmaceutical Bulletin*, 24(8), 892–895.
- 24. Okoli, C. O., Akah, P. A., & Nwafor, S. V. (2005). Anti-inflammatory activity of *Securidacalongipedunculata* root bark extracts. *Journal of Ethnopharmacology*, 102(3), 423–429.
- 25. Arul, B., Ignacimuthu, S., & Govindasamy, S. (2005). Anti-inflammatory activity of the aerial parts of *Coldenia procumbens* Linn. *Journal of Ethnopharmacology*, 96(1–2), 167–170.
- 26. Tripathi, P., Arora, S., Gupta, R., & Mali, P. R. (2011). Anti-inflammatory activity of water-soluble portion of ethanolic extract of *Nyctanthes arbor-tristis* against foreign body-



ISSN PRINT 2319 1775 Online 2320 7876

- induced inflammation. *International Research Journal of Pharmacy*, 2(3), 249–251. Available on ResearchGate
- 27. Patel, K. S., Patel, M. S., & Patel, R. J. (2011). Evaluation of anti-inflammatory activity of aqueous extracts of pericarp and seeds of *Trapa natans*. *Pharmacologyonline*, 1, 56–62. Available at Pharmacologyonline
- 28. Saha, P., Das, S., & Pal, S. (2011). Isolation and characterization of a triterpene saponin from *Bauhinia variegata* leaves with potent anti-inflammatory activity. *Journal of Natural Remedies*, 11(1), 1–6.
- 29. Yadav, R., Jain, S. K., & Sharma, R. A. (2011). Comparative evaluation of antiinflammatory activity of ethanolic and aqueous extracts of *Calllicarpa macrophylla* leaves. *Asian Journal of Pharmaceutical and Clinical Research*, 4(3), 78–81.
- 30. Mayo Clinic Proceedings. (1988). *Volume 63 Index*. Mayo Foundation for Medical Education and Research. Available via Archive.org
- 31. San doval, M., Charbonnet, R. M., Okuhama, N. N., Roberts, J., Krenova, Z., Trentacosti, A. M., & Miller, M. J. S. (2002). Cat's claw inhibits TNFα production and scavenges free radicals: Role in cytoprotection. *Free Radical Biology and Medicine*, *32*(8), 712–723.
- 32. Javed, S., Shoaib, A., & Mahmood, Z. (2005). Anti-inflammatory activity of *Colchicum luteum* and *Foeniculum vulgare* in carrageenan-induced paw edema in rats. *Pakistan Journal of Pharmaceutical Sciences*, 18(4), 45–49.
- 33. Ozbek, H. (2005). The anti-inflammatory activity of *Foeniculum vulgare* essential oil in animal models. *Phytotherapy Research*, 19(4), 310–313.
- 34. Narendhirakannan, R. T., Subramanian, S., & Kandaswamy, M. (2007). Biochemical evaluation of anti-inflammatory and anti-arthritic activity of Cleome gynandra extract in animal models. *Phytotherapy Research*, 21(6), 575–581.
- 35. Bang, J. S., Oh, D. H., Choi, H. M., Sur, B. J., Lim, S. J., Kim, J. Y., ... & Kim, D. K. (2009). Anti-inflammatory and anti-arthritic effects of piperine in human interleukin 1β-stimulated fibroblast-like synoviocytes and in rat arthritis models. *Arthritis Research & Therapy*, 11(2), R49.
- 36. Deshpande, S. S., Shah, G. B., & Parmar, N. S. (2009). Anti-inflammatory activity of *Abutilon indicum* in rats. *Indian Journal of Pharmacology*, 41(2), 87–91.
- 37. Chen, Y., Wang, Y., & Zhang, J. (2011). Anti-inflammatory and anti-arthritic activities of *Physalis angulata* leaves extract. *Journal of Ethnopharmacology*, *135*(2), 373–378.
- 38. Nair, V., Singh, S., & Gupta, Y. K. (2011). Evaluation of the disease modifying activity of *Centella asiatica* in experimental arthritis. *Journal of Ethnopharmacology*, 134(2), 434–439. https://doi.org/10.1016/j.jep.2010.12.022
- 39. Mirjalili, M. H., Moyano, E., Bonfill, M., Cusido, R. M., &Palazón, J. (2009). Steroidal lactones from *Withaniasomnifera*, an ancient plant for novel medicine. *Molecules*, 14(7), 2373–2393.
- 40. Patwardhan, B., Bodeker, G., & Vaidya, A. D. B. (2010). Ayurveda and traditional Chinese medicine: A comparative overview. *Evidence-Based Complementary and Alternative Medicine*, 2010, 1–10.
- 41. Joseph, B., & Raj, S. J. (2010). Pharmacognostic and phytochemical properties of *Aloe vera* Linn—An overview. *International Journal of Pharmaceutical Sciences Review and Research*, 4(2), 106–110.
- 42. Chopade, A. R., Sayyed, A. A., & Chaudhary, A. (2008). Green tea polyphenol epigallocatechin gallate inhibits collagen-induced arthritis in mice. *Inflammation*, 31(5), 361–368.
- 43. Akroum, S., Bendjeddou, D., & Satta, D. (2009). Antioxidant and anti-inflammatory activities of polyphenol-rich extracts from *Camellia sinensis* leaves. *Phytotherapy Research*, 23(9), 1234–1239.



ISSN PRINT 2319 1775 Online 2320 7876

- 44. Ahmed, S. (2010). Green tea polyphenol epigallocatechin-3-gallate inhibits the expression of IL-1β and TNF-α in human chondrocytes. *Journal of Nutrition*, *140*(3), 439–445.
- 45. Tripathy, S., Pradhan, D., & Anjana, M. (2010). Anti-inflammatory and antiarthritic potential of *Ammaniabaccifera* Linn. *International Journal of Pharma and Bio Sciences*, I(3), 1–10.
- 46. Subramani, J., Damodaran, A., Kanniappan, M., & Mathuram, L. N. (2009). Anti-inflammatory effect of petroleum ether extract of *Vitex negundo* leaves in rat models of acute and subacute inflammation. *Pharmaceutical Biology*, 47(4), 335–339.
- 47. Vishwanathan, A. S., &Basavaraju, R. (2010). A review on *Vitex negundo* L.—A medicinally important plant. *European Journal of Biological Sciences*, *3*(1), 30–42.
- 48. Karthikeyan, M., & Deepa, M. K. (2011). Anti-inflammatory activity of *Premnacorymbosa* (Burm.f.) Rottl. & Willd. leaves extracts in Wistar albino rats. *Asian Pacific Journal of Tropical Medicine*, 4(7), 510–513.
- 49. Prajapati, M. S., Patel, J. B., Modi, K., & Shah, M. B. (2010). *Leucas aspera*: A review. *Pharmacognosy Reviews*, 4(7), 85–87.
- 50. Kripa, K. G., Chamundeeswari, D., Thanka, J., & Reddy, C. U. M. (2010). Effect of hydroalcoholic extract of aerial parts of *Leucas aspera* on inflammatory markers in complete Freund's adjuvant-induced arthritic rats. *International Journal of Green Pharmacy*, 4(4), 281–287.
- 51. Rajendran, R., & Krishnakumar, E. (2010). Anti-arthritic activity of *Premnaserratifolia* Linn. wood against adjuvant-induced arthritis. *Avicenna Journal of Medical Biotechnology*, 2(2), 101–106.
- 52. Chakraborty, A. K., & Roy, H. K. (2010). Evaluation of anti-arthritic activity of ethanolic extract of *Cleome rutidosperma*. *Journal of Pharmaceutical Science and Technology*, 2(10), 330–332.
- 53. Otari, K. V., Bhandari, U. M., & Bhise, S. B. (2010). Evaluation of anti-arthritic activity of *Vernonia anthelmintica* Willd. seeds in experimental models. *Research and Reviews: Journal of Pharmacognosy and Phytochemistry*, 2(1), 22–28.
- 54. Amresh, G., Singh, P. N., & Rao, C. V. (2007). Antinociceptive and antiarthritic activity of *Cissampelos pareira* roots. *Journal of Ethnopharmacology*, 111(3), 531–536.
- 55. Talwar, S., Nandakumar, K., Nayak, P. G., Bansal, P., Mudgal, J., Mor, V., Rao, C. M., & Lobo, R. (2011). Anti-inflammatory activity of *Terminalia paniculata* bark extract against acute and chronic inflammation in rats. *Journal of Ethnopharmacology*, 134(2), 323–328.
- 56. Joseph, B., & Raj, S. J. (2011). Pharmacognostic and phytochemical properties of *Ficus bengalensis* Linn—An overview. *International Journal of Pharmaceutical Sciences Review and Research*, 6(2), 125–129.
- 57. Manocha, N., Chandra, S. K., Sharma, V., Sangameswaran, B., & Saluja, M. (2011). Antirheumatic and antioxidant activity of extract of stem bark of *Ficus bengalensis*. *Research Journal of Chemical Sciences*, 1(2), 2–8.
- 58. Behbahani, M., Ali, A. M., Muse, R., & Banu, N. M. M. (2007). Anti-oxidant and anti-inflammatory activities of leaves of *Barringtonia racemosa*. *Journal of Medicinal Plants Research*, *I*(1), 1–7.
- 59. Patil, K. R., Patil, C. R., Jadhav, R. B., Mahajan, V. K., Patil, P. R., & Gaikwad, P. S. (2011). Anti-arthritic activity of bartogenic acid isolated from fruits of *Barringtonia racemosa*Roxb. (Lecythidaceae). *Evidence-Based Complementary and Alternative Medicine*, 2011, 785245.
- 60. Jana, U., Chattopadhyay, R. N., & Chakraborty, A. (1999). Effect of *Tinospora cordifolia* root extract on the immune response in experimental arthritis. *Indian Journal of Pharmacology*, 31(6), 417–422.



ISSN PRINT 2319 1775 Online 2320 7876

- 61. Singh, N., & Singh, S. M. (2003). Immunomodulatory role of *Tinospora cordifolia* extract in experimental arthritis. *Indian Journal of Pharmacology*, *35*(3), 183–186. Link
- 62. Rawal, A., Singh, S. M., & Gupta, S. (2009). Modulation of immune response by *Tinospora cordifolia* in collagen-induced arthritis. *Journal of Ethnopharmacology*, 123(2), 365–372.
- 63. Paval, J., & Sumalatha, S. (2011). Evaluation of anti-arthritic activity of *Tinospora* cordifolia in experimental models. *International Journal of Applied Biology and Pharmaceutical Technology*, 2(1), 123–127.
- 64. Chandur, U., Shashidhar, S., Chandrasekar, S. B., Bhanumathy, M., & Midhun, T. (2011). Phytochemical evaluation and anti-arthritic activity of root of *Saussurealappa*. *Pharmacologia*, 2(9), 265–267.
- 65. Kimura, Y., Sumiyoshi, M., &Kawahira, K. (2011). Effects of *Tripterygium wilfordii* extract on collagen-induced arthritis in mice. *Biological and Pharmaceutical Bulletin*, 34(6), 838–844.
- 66. Saravanan, S., Babu, N. P., Pandikumar, P., &Ignacimuthu, S. (2011). Therapeutic effect of *Saracaasoca* (Roxb.) Wilde on lysosomal enzymes and collagen metabolism in adjuvant-induced arthritis. *Inflammopharmacology*, 19(4), 207–216.
- 67. Mishra, N. K., Bstia, S., Mishra, G., Chowdary, K. A., & Patra, S. (2011). Anti-arthritic activity of *Glycyrrhiza glabra*, *Boswellia serrata* and their synergistic activity in combined formulation studied in Freund's adjuvant-induced arthritic rats. *Journal of Pharmaceutical Education and Research*, 2(2), 92–98.
- 68. Khan, M. O. A., Mohiuddin, E., Usmanghani, K., Hannan, A., Akram, M., Shah, S. M. A., & Asif, M. (2011). Clinical evaluation of herbal medicines for the treatment of rheumatoid arthritis. *Pakistan Journal of Nutrition*, 10(1), 51–53.
- 69. Rehman, R., Akram, M., Naveed, A., Jabeen, Q., Saeed, T., Shah, S. M. A., & Ahmed, K. (2011). *Zingiber officinale* Roscoe: Pharmacological activity. *Journal of Medicinal Plants Research*, 5(3), 344–348.
- 70. Babu, A. R., & Karki, S. S. (2011). Anti-inflammatory activity of various extracts of roots of *Calotropis procera* against different inflammation models. *International Journal of Pharmacy and Pharmaceutical Sciences*, 3(3), 191–194.
- 71. Bhalerao, S. A., & Munjal, A. (2011). Anti-arthritic activity of *Nyctanthesarbortristis* Linn leaves. *International Journal of Pharmaceutical Sciences Review and Research*, 8(2), 123–127.
- 72. Sandhar, H. K., Kumar, B., & Sharma, P. (2011). A review on *Nyctanthesarbortristis* Linn: A medicinal plant. *International Journal of Pharmaceutical Sciences Review and Research*, 9(1), 67–71.
- 73. Paval, J., Kaitheri, S. K., Potu, B. K., Govindan, S., Kumar, R. S., Narayanan, S. N., &Moorkoth, S. (2009a). Anti-arthritic potential of the plant *Justicia gendarussa* Burm F. *Clinics*, 64(4), 357–360. https://doi.org/10.1590/S1807-59322009000400015
- 74. Paval, J., Kaitheri, S. K., Potu, B. K., Govindan, S., Kumar, R. S., Narayanan, S. N., &Moorkoth, S. (2009b). Comparing the anti-arthritic activities of the plants *Justicia gendarussa* Burm F. and *Withaniasomnifera* Linn. *International Journal of Green Pharmacy*, 3(4), 281–284. https://doi.org/10.4103/0973-8258.59732
- 75. Bachheti, R. K., Rai, I., Joshi, A., & Rana, V. (2011). Phytochemical and pharmacological profile of *Justicia gendarussa*: A review. *International Journal of Research in Pharmaceutical and Biomedical Sciences*, 2(4), 1591–1595.
- 76. Correa, M. P., & Silva, M. G. (2012). Phytochemical screening and anti-inflammatory activity of *Justicia gendarussa* extracts. *Brazilian Journal of Pharmacognosy*, 22(1), 45–50.



ISSN PRINT 2319 1775 Online 2320 7876

- 77. Rajan, S., Shaikh, M., & Mehta, A. (2012). Anti-arthritic activity of *Hemidesmus indicus* R.Br. roots in rats. *Asian Pacific Journal of Tropical Medicine*, 5(2), 130–135. https://doi.org/10.1016/S1995-7645(12)60011-X
- 78. Nair, V., Singh, S., & Gupta, Y. K. (2010). Anti-arthritic and disease modifying activity of *Terminalia chebula* Retz. in experimental models. *Journal of Pharmacy and Pharmacology*, 62(12), 1801–1806. https://doi.org/10.1111/j.2042-7158.2010.01193.x
- 79. Singh, S., & Sharma, R. (2010). Evaluation of anti-inflammatory activity of *Terminalia chebula* in experimental models. *International Journal of Pharmaceutical Sciences Review and Research*, 3(2), 123–127.
- 80. Chang, C. L., & Lin, C. S. (2012). Phytochemical composition, antioxidant activity, and neuroprotective effect of *Terminalia chebula*Retzius extracts. *Evidence-Based Complementary and Alternative Medicine*, 2012, 125247. https://doi.org/10.1155/2012/125247
- 81. Nair, V., Singh, S., & Gupta, Y. K. (2012). Evaluation of disease modifying activity of *Coriandrum sativum* in experimental models. *Indian Journal of Medical Research*, 135, 240–245. Link to article
- 82. Patel, R. G., Pathak, N. L., Rathod, J. D., Patel, L. D., & Bhatt, N. M. (2012). Phytopharmacological properties of *Randia dumetorum* as a potential medicinal tree: An overview. *Journal of Applied Pharmaceutical Science*, 2(3), 36–40. PDF link
- 83. Mazumder, P. M., Arulmozhi, S., Mondal, A., Rathinavelusamy, P., &Sasmal, D. (2012). Evaluation of antiarthritic and immunomodulatory activity of *Barlerialupulina*. *Asian Pacific Journal of Tropical Biomedicine*, 2(3, Supplement), S1402–S1407. https://doi.org/10.1016/S2221-1691(12)60425-0
- 84. Thirumal, M., Vijaya Bharathi, R., Kumudhaveni, B., & Kishore, G. (2013). Anti-arthritic activity of chloroform extract of *Barringtonia acutangula* (L.) Gaertn. leaves on Wistar rats. *Der Pharmacia Lettre*, 5(3), 367–373. PDF link
- 85. Mishra, N. K., Biswal, G. S., Chowdary, K. A., & Mishra, G. (2013). Anti-arthritic activity of *Tribulus terrestris* studied in Freund's adjuvant-induced arthritic rats. *Journal of Pharmaceutical Education and Research*, 4(1), 41–46. PDF link
- 86. Rajaram, C., Reddy, K. R., & Chandra Sekhar, K. B. (2015). Evaluation of anti-arthritic activity of *Caesalpinia pulcherrima* in Freund's complete adjuvant-induced arthritic rat model. *Journal of Young Pharmacists*, 7(2), 65–70. https://doi.org/10.5530/jyp.2015.2.12

